

# Effect of the β-Adrenoceptor Agonist Flerobuterol on Serotonin Synthesis in the Rat Brain

Ko Tsuiki,\* Pierre Blier† and Mirko Diksic\*‡

\*Cone Laboratory for Neurosurgical Research, Department of Neurology and Neurosurgery, and Montreal Neurological Institute, and †Department of Psychiatry, McGill University, Montreal, PQ H3A 2B4, Canada

**ABSTRACT.** The influence of 2- and 14-day treatments with flerobuterol, a preferential  $\beta_2$ -adrenoceptor agonist, on regional serotonin (5-HT) synthesis in the rat brain was studied by autoradiography using  $\alpha$ -[\$^14C]methyl-L-tryptophan. Flerobuterol was delivered at a rate of 0.5 mg/kg/day using osmotic pumps implanted s.c. The 2-day flerobuterol treatment significantly increased plasma Trp, both free and total, and decreased plasma Leu and Ile. This resulted in a significant increase in the facilitated transport of Trp. There was a significant increase in the synthesis of 5-HT in the 2-day treatment group in the dorsal and median raphe as well as in all postsynaptic structures, with the exception of the hypothalamus. In contrast, after a 14-day treatment, the enhanced facilitated transport of Trp was no longer present, and the increase in the rate of 5-HT synthesis persisted only in the parietal and occipital cortex and the superior colliculus. These data suggest that flerobuterol, similar to other \$\beta\$-adrenergic agonists, acutely increases 5-HT synthesis, in part, through an elevation of brain Trp availability.

BIOCHEM PHARMACOL **59**;6:673–679, 2000. © 2000 Elsevier Science Inc.

KEY WORDS. α-methyl-L-tryptophan; depression; serotonin synthesis; chronic and acute treatments

It was reported that β-adrenoceptor agonists, such as isoprenaline, can increase the brain concentration of L-Tyr and L-Trp, while the plasma concentration of many amino acids is decreased [1]. This suggests that brain uptake, at least of some amino acids, can be controlled, in part, by  $\beta$ -adrenergic receptors. The free fraction of Trp in the plasma, however, was not measured concomitantly in these studies. The increase of brain Trp could have special relevance with respect to the synthesis of 5-HT\s because the enzyme tryptophan hydroxylase, the rate-limiting enzyme in the synthesis of 5-HT, is not saturated by Trp. Flerobuterol, a preferential  $\beta_2$ -adrenoceptor agonist ( $K_i$  = 926 and 518 nM for  $\beta_1$ - and  $\beta_2$ -adrenoceptor binding, respectively [2]), exhibits antidepressant activity in animals and was shown to enhance serotonergic neurotransmission using in vivo electrophysiological techniques [3, 4]. Furthermore, the  $\beta_2$ -adrenergic agonist clenbuterol is reported to act as an antidepressant in depressed patients [5]. It is believed that enhanced 5-HT neurotransmission in some situations is directly related to the 5-HT concentration, such as during MAO inhibition, and it probably is also dependent on the rate of 5-HT synthesis. Since the major classes of antidepressant treatment, including MAO inhibitors, enhance 5-HT neurotransmission [6], it is thus pos-

The influence of norepinephrine on the brain serotonergic system has been documented in different types of investigations [7–9]. It has been shown that the brain uptake of the LNAA (e.g. Trp, Tyr) can be affected by the β-adrenergic agonists isoprenaline [10], isoproterenol, salbutamol [11], and clenbuterol [12] despite a significant reduction in a majority of the LNAA in the rat plasma. These investigations did not determine the fraction of free Trp in the plasma and, thus, it cannot be concluded that the influence of the Trp free fraction is related to the increase in the brain Trp. The conclusion was that, despite a reduction in the plasma concentration of the majority of the LNAA, their brain concentrations were increased. This might suggest that the transport of LNAA is under the direct control of β-adrenergic receptors. Because of these data, suggesting a direct effect of  $\beta$ -adrenergic agonists, we have measured plasma concentration of the LNAA, and calculated the facilitated Trp flux ( $\nu$ ) separately for total  $(\nu_{\rm T})$  and free  $(\nu_{\rm f})$  plasma Trp [13]. These quantities would reflect facilitated flux of Trp in competition by other LNAA sharing the same carrier.

In the present study, measurements of the regional 5-HT synthesis using a recently developed autoradiographic method were carried out [14–16]. It was deemed important to investigate brain 5-HT synthesis in rats treated with flerobuterol for different lengths of time because it was shown that short- and long-term flerobuterol administra-

sible that flerobuterol could act on the 5-HT system and exert its antidepressant-like effect in animals by altering 5-HT synthesis.

<sup>‡</sup> Corresponding author: Mirko Diksic, Ph.D., Montreal Neurological Institute, 3801 University St., Montreal, QC H3A 2B4, Canada. Tel. (514) 398-8526; FAX (514) 398-8195.

<sup>§</sup> Abbreviations: 5-HT, serotonin; α-MTrp, α-methyl-L-tryptophan; MAO, monoamine oxidase; and LNAA, large neutral amino acids.

Received 24 February 1999; accepted 10 August 1999.

674 K. Tsuiki et al.

tion differentially affect the brain 5-HT system [4]. The  $\alpha$ -MTrp method utilizes tracer kinetics and does not require the administration of any other drug to measure the rate of synthesis, thus precluding any influence of auxiliary drugs such as the aromatic amino acid decarboxylase inhibitor NSD-1015, which can, on its own, alter 5-HT synthesis [17].

# MATERIALS AND METHODS Materials

 $\alpha$ -[<sup>14</sup>C]MTrp (sp. act.  $\approx 55$  mCi/mmol) was synthesized by us, with the method described by Mzengeza et al. [18], from [<sup>14</sup>C]CH<sub>3</sub>I purchased from Amersham Canada Ltd. The stereochemical purity was confirmed by HPLC using a chiral column. The chiral and radiochemical purity of the radiopharmaceutical was greater than 98%. Ultrafree-MC filters (Cat. No. UF3LGCOO) with a 10,000 MW cutoff point were purchased from Millipore Canada Ltd. The HPLC solvents were purchased from the Baker Chemical Co.

# **Experimental Procedures**

Male Sprague-Dawley rats weighing about 250 g were treated continuously for 2 or 14 days with saline (controls) or with flerobuterol (0.5 mg/kg/day, s.c.), delivered by osmotic minipump (pump models 2ML1 and 2ML2 for 2and 14-day infusion, respectively). The minipumps were implanted under chloral hydrate anaesthesia (400 mg/kg, i.p.). This dose of flerobuterol was chosen as the one producing central effects with minimal peripheral effects [3]. After implantation of minipumps, rats were returned to their cages. After either 2 or 14 days of treatment, the rats were anaesthetized with halothane (1 to 1.5%), and arterial and venous catheters were implanted, as part of a normal tracer procedure (see details for animal handling in Ref. 15). Animals were food-deprived for some 20 hr before injection of tracer; however, water was provided ad lib. About 2 hr after the rats woke up from anaesthesia, about 50  $\mu$ Ci of  $\alpha$ -[14C]MTrp in 2 mL of saline was injected over 2 min using an infusion pump. The arterial plasma samples were taken at increased time intervals for input function determination, and additional samples were collected for the determination of the plasma amino acids (Val, Leu, Ile, Met, Tyr, Phe, Trp) and the plasma concentration of free Trp. The free Trp was determined in the plasma ultrafiltrate [14]. Arterial pCO<sub>2</sub>, pO<sub>2</sub>, pH, blood pressure, and hematocrit were checked periodically, and in all animals were within laboratory standards. Body temperature, measured through a rectal probe, was maintained at 37° with a heating lamp. To minimize as much as possible any influence of circadian rhythm on the results, the tracer was always injected between noon and 2:00 p.m. Rats were decapitated 60 or 150 min after tracer injection, and brains were extracted, frozen, and cut at about  $-20^{\circ}$  into 30  $\mu$ m slices in a microtome. The brain slices were contacted to an x-ray sensitive film for 3 weeks along with <sup>14</sup>C-standards (tissue equivalent) and developed. Quantification of autoradiograms was done with the aid of a computerized image analyzer (The Image Calculator, Soquelec Ltd.), and tracer concentrations (nCi/g tissue) were determined in thirty structures.

# Calculation of the Rate of 5-HT Synthesis

The rates of 5-HT synthesis were calculated by using an approximation method described in details in our previous publications [14, 16, 19]. The method was shown to give results not significantly different from those obtained by the two-time point method described by Nagahiro et al. [15]. The method is based on the assumption that the apparent volume of distribution of the precursor pool (V<sub>0</sub>; mL/g) is approximately the same in all brain structures, and has a value of  $0.45 \pm 0.1$  mL/g [16, 19]. In the calculations presented here, equality of the Vo in the control and treatment groups was assumed. Tissue tracer concentrations in each structure and for individual rats were converted into volume of distribution (V<sub>D</sub>) by dividing it with the plasma tracer concentration at the end of the experiment  $[C_p^*(T)]$ . The  $V_D$  was then corrected for the contribution of the precursor pool by subtracting 0.45 (Vo; mL/g) and dividing by the exposure time  $\Theta \left[\Theta = \int_0^T C_p^*(t) dt / C_p^*(T) \right]$ . The rates of 5-HT synthesis were calculated as per Eq. 1 for each structure and for individual rats, and the average for the group and the standard deviations were calculated from these individual values [16, 19],

$$R = \frac{C_p}{IC} \cdot \frac{V_D - V_0}{\Theta} \tag{1}$$

where  $C_p$  (pmol/mL) is the plasma concentration of free Trp and LC is the conversion factor taken to be equal to 0.42  $\pm$  0.07 [20]. LC is a conversion factor that converts the rate of tissue uptake of tracer ( $\alpha$ -[1<sup>4</sup>C]MTrp) into the uptake of Trp via 5-HT pathway, which after multiplying by plasma free Trp gives the rate of 5-HT synthesis [20].

The facilitated Trp flux ( $\nu$ ; nmol/g/min) from blood to brain was calculated by the Michaelis–Menten formula [21],

$$\nu = \frac{C_p \cdot V_{\text{max}}^{Trp}}{K_m^{Trp} \left(1 + \frac{C_p^{AA}}{K_m^{AA}}\right) + C_p}$$
 (2)

where  $C_p$  (nmol/mL) is the plasma concentration of Trp, and  $C^{AA}$  and  $K_m^{AA}$  are concentrations and the Michaelis–Menten constant, respectively for the competing amino acids. The  $K_m$  values used in these calculations were taken from Miller *et al.* [22].

The HPLC system consisted of a reverse phase column, a fluorescent detector (excitation filter, 330 nm; fluorescence filter, 465 nm), and the buffer as elution solvent with post-column derivatization. Standards were prepared from

# Flerobuterol 2 day Experiment AD ED PB ID MD DR DR DR DR ND VI

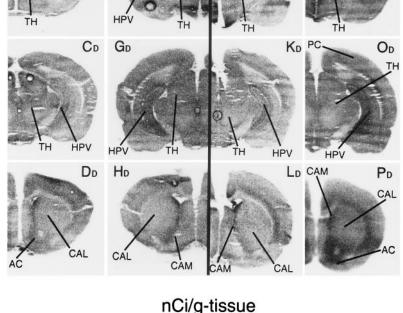


FIG. 1. A set of radiochromatograms obtained in rats treated with flerobuterol (0.5 mg/kg/day delivered by osmotic pump) for 2 days (images  $I_D$  to  $P_D$ ) and respective controls receiving saline without flerobuterol (images  $A_D$  to  $H_D$ ). Images given in  $A_D$  to  $D_D$  and  $I_D$  to  $L_D$  are for rats killed 60 min after tracer injection, and those given in  $E_D$  to  $H_D$  and  $H_D$  to  $H_D$  are for rats killed 150 min after tracer

injection. Some brain structures are identified with letters: dorsal raphe (DR); pineal body (PB); hippocampus-dorsal (HPD); hippocampus-ventral (HPV); thalamus (TH); caudate-lateral (CAL); caudate-medial (CAM); nucleus accumbens (AC); parietal

87.2

151.8

pure chemical and contained about 20 nmol/mL of each LNAA.

cortex (PC); and layer VI of cortex (VI).

1.4

13.5

39.4

## **RESULTS**

There were no significant differences in the physiological parameters or body weight between the groups of control and treated rats. A representative set of autoradiograms is shown in Fig. 1 for 2-day control and treated rats, and in Fig. 2 for 14-day control and treated rats. In general, a large accumulation of the labelled tracer in the brain structures

known to have a large concentration of serotonergic cells (e.g. raphe) was readily detectable.

335.0

169.8 249.8

Plasma concentrations of Trp and other amino acids are given in Table 1. There was a decrease in 2-day flerobuterol-treated rats in the plasma concentration of Ile (24%) and Leu (21%) and an increase of total and free plasma Trp (54 and 57%, respectively). In contrast, in the 14-day experiments, there was only a significant increase of the Leu concentration as compared with the respective controls. In the 2-day group, there was also an increase in the facilitated Trp flux, calculated with both free ( $\nu_f$ ) and

676 K. Tsuiki et al.

# Flerobuterol 2 Week Experiment

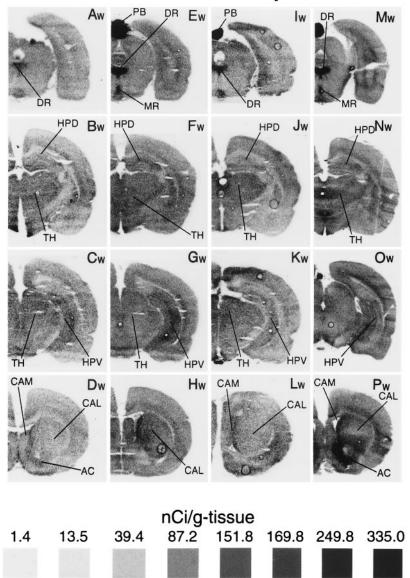


FIG. 2. A set of radiochromatograms obtained in rats treated with flerobuterol (0.5 mg/kg/day delivered by osmotic pump) for 14 days (images  $I_W$  to  $P_W$ ) and the respective controls receiving saline without flerobuterol (images  $A_W$  to  $H_W$ ). Images given in  $A_W$  to  $D_W$  and  $I_W$  to  $L_W$  are for rats killed 60 min after tracer injection, and those given in  $E_W$  to  $E_W$  and  $E_W$  are for rats killed 150 min after tracer injection. Some brain structures are identified with letters: dorsal raphe (DR); median raphe (MR); pineal body (PB); hippocampus-dorsal (HPD); hippocampus-ventral (HPV); thalamus (TH); caudate-lateral (CAL); caudate-medial (CAM); and nucleus accumbens (AC).

total ( $\nu_{\rm T}$ ) Trp. The increase could be related, in part, to a significant increase of plasma Trp and to a decrease in Leu and Ile. Since there is no evidence that plasma Trp is linearly related to the brain 5-HT synthesis rate, it would not be appropriate to normalize brain 5-HT synthesis rates according to plasma Trp. In short, the change in the plasma Trp and some other amino acids should be considered as characteristics of the flerobuterol treatment.

The rates of synthesis in selected discrete brain structures are given in Table 2 for 2-day flerobuterol-treated rats and in Table 3 for 14-day flerobuterol-treated rats. In the 2-day treatment group, a highly significant increase,

as compared with the 2-day controls, was observed in the rate of 5-HT synthesis. In the 14-day experiment, only parietal and occipital cortices and the superior colliculus still showed an increase in 5-HT synthesis, while there was a significant decrease in the mammillary body and the medial part of the caudate nucleus (Table 3). It should be noticed that while 2-day treatments produced decreases in the plasma concentrations of some amino acids with a significant increase in total and free Trp, the 14-day treatment resulted in a significant increase (15-%) in leucine (Table 1) with non-significant changes in other amino acids.

TABLE 1. Plasma concentration of some essential amino acids sharing the same carrier with Trp in 2- and 14-day flerobuterol-treated rats and respective controls

	2 days		14 days	
	Control (N = 10)	Treated (N = 10)	Controls (N = 14)	Treated (N = 13)
	P	lasma concentration (nmol/mL)	1	
Val	$364 \pm 45$	$324 \pm 67$	$406 \pm 34$	$424 \pm 31$
Met	$66 \pm 7$	$64 \pm 7$	$66 \pm 5$	$61 \pm 6$
Lle	$167 \pm 21$	$127 \pm 30*$	$97 \pm 11$	$106 \pm 15$
Leu	$337 \pm 37$	$265 \pm 35*$	$254 \pm 21$	$291 \pm 19*$
Tyr	$65 \pm 7$	$66 \pm 9$	$94 \pm 8$	$94 \pm 9$
Phe	$104 \pm 15$	99 ± 14	$99 \pm 8$	$109 \pm 13$
Trp (total)	$44 \pm 10$	$68 \pm 15*$	$82 \pm 10$	$89 \pm 10$
Trp (free)	$3.7 \pm 0.6$	$5.8 \pm 0.9*$	$6.6 \pm 1.5$	$7.2 \pm 1.7$
$v_f^{\dagger}$	$0.098 \pm 0.027$	$0.17 \pm 0.04$	$0.19 \pm 0.07$	$0.19 \pm 0.07$
$v_T^{\dagger}$	$1.10 \pm 0.37$	$1.84 \pm 0.56*$	$2.09 \pm 0.60$	$2.12 \pm 0.57$

Values are means ± SD; N represents the number of rats.

# **DISCUSSION**

In the present study, the 2-day flerobuterol treatment resulted in a decrease of some amino acids in plasma, but in a significant increase of plasma Trp (Table 1). This decrease of some amino acids and increase in the free Trp resulted in a significant increase of the facilitated flux of Trp (both  $\nu_f$  and  $\nu_T$ ). In addition to the facilitated transport of Trp into the brain, there could be a contribution from diffusion of Trp because the diffusion constant in normal rat brain seems to be substantial [23]. This increase in the influx of Trp (both facilitated and diffusion) is responsible, at least in part, for the increases of Trp reported in the brain

TABLE 2. Comparison of the rates of 5-HT synthesis in saline-treated (control) rats with that in rats injected for 2 days with flerobuterol at a rate of 0.5 mg/kg/day

Structure	Control	Treated	% Difference from controls*
5-I	TT synthesis	(pmol/g/min)	
Dorsal raphe	$104 \pm 12$	$130 \pm 12$	+25
Medial raphe	$87 \pm 12$	$116 \pm 11$	+33
Hippocam. ventral	$58 \pm 6$	$83 \pm 13$	+43
Hippocam. dorsal	$51 \pm 4$	$72 \pm 13$	+41
Hypothalamus	$55 \pm 9$	$57 \pm 11$	+9 (NS)
Thalamus	$41 \pm 4$	$63 \pm 10$	+50
Mamillary body	$46 \pm 7$	$70 \pm 12$	+52
Ventr. tegm. area	$43 \pm 5$	$58 \pm 12$	+35
Parietal cortex	$39 \pm 5$	$53 \pm 9$	+36
Temporal cortex	$47 \pm 7$	$65 \pm 14$	+38
Occipital cortex	$39 \pm 4$	$56 \pm 7$	+44
Caudate-medial	$51 \pm 7$	$69 \pm 13$	+35
Caudate-lateral	$50 \pm 8$	$65 \pm 11$	+30
Medial geniculate	$54 \pm 6$	$80 \pm 13$	+48
Superior colliculus	$54 \pm 4$	$76 \pm 12$	+41
Amygdala	$42 \pm 4$	$56 \pm 13$	+33

Values are means ± SD of ten rats.

following treatments with adrenergic agonists [10–12]. It is of interest to note that in 14-day flerobuterol-treated rats there was no significant difference in the plasma free or total Trp between control and treated rats (Table 1). In addition, there was only a significant increase in leucine in the 14-day treated group. However, there was no significant differences in the facilitated transports of either total or free Trp ( $\nu_f$  and  $\nu_T$ ; Table 1).

In the autoradiographic experiments, a significant increase in 5-HT synthesis was observed in all brain structures except for the hypothalamus in the 2-day treatment group

TABLE 3. Comparison of the rates of 5-HT synthesis in saline-treated (control) rats with that in rats injected for 14 days with a continuous infusion of 0.5 mg/kg/day of flerobuterol

Structure	Control $(N = 14)$	Treated (N = 13)	% Difference from controls*				
5-HT synthesis (pmol/g/min)							
Dorsal raphe	$178 \pm 16$	$187 \pm 16$	NS				
Medial raphe	$135 \pm 13$	$131 \pm 12$	NS				
Hippocam. ventral	$73 \pm 7$	$70 \pm 9$	NS				
Hippocam. dorsal	$62 \pm 7$	$56 \pm 10$	NS				
Hypothalamus	$74 \pm 7$	$72 \pm 7$	NS				
Thalamus	$46 \pm 4$	$42 \pm 7$	NS				
Mamillary body	$196 \pm 15$	$172 \pm 20$	$-14\dagger$				
Ventr. tegm. area	$112 \pm 11$	$110 \pm 18$	NS				
Parietal cortex	$38 \pm 7$	$46 \pm 7$	+20†				
Temporal cortex	$61 \pm 7$	$62 \pm 9$	NS				
Occipital cortex	$49 \pm 6$	$54 \pm 7$	+10†				
Caudate-medial	$74 \pm 12$	$62 \pm 17$	$-16\dagger$				
Caudate-lateral	$63 \pm 14$	$60 \pm 15$	NS				
Medial geniculate	$59 \pm 7$	$62 \pm 9$	NS				
Superior colliculus	$58 \pm 6$	$66 \pm 10$	+13†				
Amygdala	$67 \pm 10$	$65 \pm 13$	NS				

Values are means ± SD; N represents the number of rats.

<sup>\*</sup>Indicates a significant difference (p < 0.05; ANOVA).

<sup>†</sup>Represent-facilitated Trp fluxes calculated according to Eq. 2.

<sup>\*</sup>All differences were significant (P < 0.05; ANOVA) except that in hypothalamus, indicated as NS.

<sup>\*</sup>All differences were determined using ANOVA.

<sup>†</sup>Indicates a significant (P < 0.05) difference when compared with the control group.

678 K. Tsuiki et al.

(Table 2). Since there was a similar increase in plasma free Trp, and the plasma free Trp has been related by many investigations to brain 5-HT synthesis [13, 24], the increase in the brain 5-HT synthesis could be attributed, in large part, to the increase in the plasma Trp. It also should be noted (see above) that Trp has a rather large passive diffusion component [23] that also could contribute Trp to the brain Trp pool. It is of interest to note that the increase in the raphe nuclei is similar to the increase in other structures, suggesting that there was no additional influence on 5-HT synthesis mediated via somatodendritic 5-HT<sub>1A</sub> autoreceptors, which are known to reduce cell body 5-HT synthesis after an increase in the tissue 5-HT levels [17, 25, 26]. In our previous work, the 5-HT releaser/reuptake blocker fenfluramine reduced synthesis in the cell body region of 5-HT neurons [26], while the 5-HT<sub>1A</sub> agonist buspirone acutely decreased 5-HT synthesis throughout the rat brain [27]. However, one has to consider that fenfluramine also decreases 5-HT neuronal firing as it enhances the extracellular availability of 5-HT [28], which would, in turn, account for the decreased 5-HT synthesis rate. In this respect, the reduction in the firing of dorsal raphe neurons reported by Bouthillier et al. [4] in 2-day flerobuteroltreated rats would support results indicating an increase in the 5-HT synthesis in the 2-day treated rats, because by increasing 5-HT synthesis, flerobuterol likely increases the availability of 5-HT. This interpretation finds direct support by the restoration of 5-HT neuronal firing in 2-day flerobuterol-treated rats using the 5-HT<sub>1A</sub> autoreceptor antagonist spiperone [4]. Our observation that flerobuterol increases the rate of 5-HT synthesis is also in agreement with previous reports that β-adrenoceptor agonists increase 5-HT synthesis in the rat brain [29–31].

In the 14-day flerobuterol treatment, there remained a significant increase in 5-HT synthesis in only three structures (Table 3). Since there was no difference in the plasma Trp between treated and control groups (Table 1), the changes in the 5-HT synthesis observed could be directly related to pharmacological action(s) of flerobuterol. In 14-day flerobuterol-treated rats, Bouthillier et al. [4] reported a significantly increased effect of electrical stimulation of the ascending 5-HT in suppressing the firing activity of dorsal hippocampus pyramidal neurons. This was suggested to be the result of a greater amount of 5-HT released for each stimulation-triggered action potential. However, the synthesis measurement in the dorsal hippocampus reported here showed no significant alteration in the 5-HT synthesis rate in the flerobuterol-treated rats when compared with the respective controls. Since Bouthillier et al. [4] reported an enhanced 5-HT neurotransmission in the dorsal hippocampus in 14-day flerobuterol-treated rats, the present data suggest that this enhanced neurotransmission is not correlated with 5-HT synthesis unless there is inhibition of MAO with flerobuterol or its metabolites, which has not been shown thus far. Therefore, the exact neurobiological mechanism by which flerobuterol enhances 5-HT neurotransmission in the rat hippocampus following a 2-week treatment remains to be elucidated.

The effect of flerobuterol on 5-HT synthesis observed after 2 days but not 14 days of treatment was likely due to adaptive processes in the 5-HT rather than the  $\beta$ -adrenergic system. Indeed, there is little physiological evidence for adaptation of  $\beta_2$ -adrenoceptors following their long-term activation. Patients with asthma who inhale salbutamol several times a day do not present any clear evidence of attenuated responsiveness, certainly not over a time period similar to that used in the present study.

In summary, short-term flerobuterol treatment has a significant enhancing effect on plasma Trp and some other amino acids competing for the same blood–brain barrier transporter. A 2-day treatment produced a significant increase in 5-HT synthesis throughout the brain, which persisted in only a few structures as the treatment was prolonged to a 14-day period. This adaptive change, and the gradual enhancement of 5-HT neurotransmission observed in a previous electrophysiological study [4], are fully consistent with the delayed onset of action of  $\beta_2$ -adrenoceptor agonists in major depression.

The research reported here was supported, in part, by the MRC (MT-13368) and the U.S. Public Health Service (RO1-NS-29629)

### References

- Eriksson T and Carksson A, β-Adrenergic control of brain uptake of large neutral amino acids. Life Sci 42: 1583–1589, 1988.
- Zini R, Morin D and Tillement JP, Interaction of flerobuterol, a new antidepressant drug with beta adrenoceptors in rat CNS. Psychopharmacology Suppl 96: 276, 1988.
- 3. Puech AJ, Henry M, Martin P, Thivenet G, Rambert FA and Duteil J, Preferential central versus peripheral effect of the beta agonist flerobuterol in rats: Comparison with salbutamol. *Psychopharmacology Suppl* **96:** 276, 1988.
- Bouthillier A, Blier P and de Montigny C, Flerobuterol, a β-adrenoceptor agonist, enhances serotonergic neurotransmission: An electrophysiological study in the rat brain. Psychopharmacology 103: 357–365, 1991.
- Simon P, Lecrubier Y, Jouvent R, Puech A and Widlöcher D, Beta-receptor stimulation in the treatment of depression. Adv Biochem Pharmacol 39: 293–300, 1984.
- Haddjeri N, Blier P and de Montigny C, Long-term antidepressant treatments result in a tonic activation of forebrain 5-HT<sub>1A</sub> receptors. *J Neurosci* 18: 10150–10156, 1998.
- Trulson ME and Jacobs BL, Raphe unit activity in freely moving cats: Correlation with level of behavioral arousal. Brain Res 163: 135–150, 1979.
- Clement HW, Gemsa D and Wesemann W, The effect of adrenergic drugs on serotonin metabolism in the nucleus raphe dorsalis of the rat, studied by *in vivo* voltammetry. *Eur J Pharmacol* 217: 43–48, 1992.
- Maura G, Gemignani A and Raiteri M, Noradrenaline inhibits central serotonin release through α<sub>2</sub>-adrenoceptors located on serotonergic nerve terminals. Naunyn Schmiedebergs Arch Pharmacol 320: 272–274, 1982.
- Christensen HN, Recognition sites for material transport and information transfer. In: Current Topics in Membranes and

- Transport (Eds. Bonner F and Kleinzeller A), pp. 227–258. Academic Press, New York, 1975.
- Takao Y, Kamisak Y and Itoh T, β-Adrenergic regulation of amine precursor amino acid transport across the blood-brain barrier. Eur J Pharmacol 215: 245–251, 1992.
- 12. Edwards DJ, Sorisio DA and Knopf S, Effects of the  $\beta_2$ -adrenoceptor agonist clenbuterol on tyrosine and tryptophan in plasma and brain of the rat. Biochem Pharmacol 38: 2957–2965, 1989.
- Bloxam DL, Tricklebank MD, Patek AJ and Curzon G, Effects of albumin, amino acids, and clofibrate on the uptake of tryptophan by the rat brain. J Neurochem 34: 43–49, 1980.
- 14. Diksic M, Nagahiro S, Sourkes TL and Yamamoto YL, A new method to measure brain serotonin synthesis *in vivo*. I. Theory and basic data for a biological model. *J Cereb Blood Flow Metab* 10: 1–12, 1990.
- Nagahiro S, Takada A, Diksic M, Sourkes TL, Missala K and Yamamoto YL, A new method to measure brain serotonin synthesis in vivo. II. A practical autoradiographic method tested in normal and lithium-treated rats. J Cereb Blood Flow Metab 10: 13–21, 1990.
- Diksic M, Nagahiro S and Grdisa M, The regional rate of serotonin synthesis estimated by the α-methyl-tryptophan method in rat brain from a single time point. J Cereb Blood Flow Metab 15: 806–813, 1995.
- 17. Mück-Seler D and Diksic M, The acute effects of reserpine and NSD-1015 on the brain serotonin synthesis rate measured by an autoradiographic method. *Neuropsychopharmacology* 12: 251–262, 1995.
- Mzengeza S, Venkatachalam TK, Rajagopal S and Diksic M, Synthesis of enantiomerically pure α-[<sup>14</sup>C]methyl-L-tryptophan. Appl Radiat Isot 44: 1167–1172, 1993.
- Tsuiki K, Mück-Seler D and Diksic M, Autoradiographic evaluation of the influence of hypothalamic 5,7-dihydroxytryptamine lesion on brain serotonin synthesis. *Biochem Pharmacol* 49: 633–642, 1995.
- Vanier M, Tsuiki K, Grdisa M, Worsley K and Diksic M, Determination of the lumped constant for the α-methyltryptophan method of estimating the rate of serotonin synthesis. J Neurochem 64: 624–635, 1995.

- 21. Dixon M, The determination of enzyme inhibitor constants. *Biochem J* **55:** 153–162, 1953.
- 22. Miller LP, Pardridge WM, Braun LD and Oldendorf WH, Kinetic constants for blood-brain barrier amino acid transport in conscious rats. *J Neurochem* **45:** 1427–1432, 1985.
- Takada A, Grdisa M, Diksic M, Gjedde A and Yamamoto YL, Rapid steady-state analysis of blood-brain transfer of L-Trp in rat, with special reference to the plasma protein binding. Neurochem Int 23: 351–359, 1993.
- 24. Salter M, Knowles RG and Pogson CI, How does displacement of albumin-bound tryptophan cause sustained increases in the free tryptophan concentration in plasma and 5-hydroxytryptamine synthesis in the brain? Biochem J 262: 365–368, 1989.
- 25. Mück-Seler D and Diksic M, DL-Fenfluramine increases the 5-HT synthesis rate in the terminals while decreasing it in the cell bodies of the rat brain. *Brain Res* **737**: 45–50, 1996.
- Mück-Seler D, Jevric-Causevic A, and Diksic M, Influence of fluoxetine on regional serotonin synthesis in the rat brain. J Neurochem 67: 2434–2442, 1996.
- Okazawa H, Yamane F, Blier P and Diksic M, Effects of acute and chronic administration of the serotonin<sub>1A</sub> agonist buspirone on serotonin synthesis in the rat brain. *J Neurochem* 72: 2022–2031, 1999.
- 28. Scuvée-Moreau J and Dresse A, Influence of fenfluramine and norfenfluramine stereoisomers on the firing rate of central monoaminergic neurons in the rat. *Eur J Pharmacol* 179: 211–215, 1990.
- Erdö SL, Kiss B and Rosdy B, Effect of salbutamol on the cerebral levels, uptake and turnover of serotonin. Eur J Pharmacol 78: 357–361, 1982.
- Ningaonkar VL, Green AR, Cowen PJ, Heal DJ, Grahame-Smith DG and Deakin JFW, Studies on the mechanisms by which clenbuterol, a β-adrenoceptor agonist, enhances 5-HTmediated behaviour and increases metabolism of 5-HT in the brain of the rat. Neuropharmacology 22: 739–749, 1983.
- Waldmeier PC, Stimulation of central serotonin turnover by β-adrenoceptor agonists. Naunyn Schmiedebergs Arch Pharmacol 317: 115–119, 1981.